A kinetic study of roadside grass pyrolysis and digestate from anaerobic mono-digestion

- 3 Robert Bedoić^a, Vesna Ocelić Bulatović^b, Lidija Čuček^c, Boris Ćosić^a, Ana Špehar^d,
- 4 Tomislav Pukšec^a, Neven Duić^a
- 5

6 ^aUniversity of Zagreb, Faculty of Mechanical Engineering and Naval Architecture, Ivana Lučića 5, Zagreb, Croatia

- 7 ^bUniversity of Zagreb, Faculty of Metallurgy, Aleja narodnih heroja 3, Sisak, Croatia
- 8 ^cUniversity of Maribor, Faculty of Chemistry and Chemical Engineering, Smetanova ulica 17, Maribor, Slovenia
- 9 ^dAgroproteinka d.d. Strojarska cesta 11, Sesvete, Croatia

10 Abstract

11 The aim of this research is to evaluate the thermogravimetric behaviour of roadside grass and its digestate 12 obtained from mesophilic anaerobic mono-digestion by quantifying its impacts on biomass composition 13 and properties. Thermogravimetric measurements were conducted in a laboratory furnace under nitrogen 14 flowrate of 100 mL/min in the temperature range from 35 to 800°C at five different heating rates of 2.5, 5, 15 10, 15 and 20°C/min. Friedman and Kissinger-Akahira-Sunose differential and integral isoconversional 16 models were applied to determine the distributions of activation energies and modified pre-exponential 17 factors per reacted mass (degree of conversion). The investigation demonstrated that anaerobic digestion 18 of roadside grass can be used to generate biochar-richer material (with significantly greater yield of final 19 residues after pyrolysis) with less energy required for subsequent pyrolysis in comparison with raw 20 roadside grass.

21

22 Keywords:

Roadside grass; roadside grass digestate; anaerobic digestion; pyrolysis; kinetic study; thermogravimetric
 measurements; isoconversional models

25

26 1. Introduction

Grass is a sustainable source of lignocellulosic material which can be cultivated on non-arable
lands, making it non-competitive with other crops for food production (Rodriguez et al., 2017). The
location of the area where grass is collected impacts its chemical composition and further application.
Grass collected on non-arable lands has been shown to be an abundant and environment-friendly source of

31 material to produce bioenergy in the form of biogas (Bedoić et al., 2019). Another efficient and

32 environmentally attractive process for converting biomass and different types of waste to energy products

33 is pyrolysis (Campuzano et al., 2019). Besides anaerobic digestion and pyrolysis, biomass can be

34 converted to energy and bioproducts through combustion, gasification and biochemical processing

35 (Akhtar et al., 2018).

Pyrolysis is one of the most thoroughly researched thermochemical conversion processes of biomass into valuable hydrocarbon and alternative fuels (Dhyani and Bhaskar, 2018). Slow pyrolysis producing charcoal has been successfully utilised for waste-to-energy and waste-to-liquid purposes (Rostek and Biernat, 2013). Pyrolysis can be studied under different atmospheric conditions (inert or reactive) that affect the complexity of the thermal behaviour of feedstock (Mikulcic et al., 2019). Gaseous products that appear during pyrolysis contain variable chemical constituents based on the feedstock used (Stančin et al., 2019).

Pyrolysis of biomass and waste is widely examined with thermogravimetric analysis (TGA) together with kinetic studies (Zhang et al., 2017). To evaluate the effects of different waste materials on pyrolysis, various experiments have been performed using the TGA (Oyedun et al., 2014). Thermochemical processes (pyrolysis, gasification and combustion) of different types of lignocellulosic biomass have shown that for describing their kinetics, different approaches/models are suggested (Senneca, 2007). Generally, there are two types of kinetic methods which have been used for the description of biomass and waste pyrolysis kinetics, model-fitting and isoconversional or model-free methods (Jain et al., 2016).

50 Isoconversional methods are more popular in the analysis of biomass pyrolysis kinetics than model-

51 fitting methods (Burnham and Dinh, 2007). The advantages of isoconversional methods include

52 computation of kinetic parameters without modelling assumptions (Ramajo-Escalera et al., 2006) and

53 better suitability for more complex and multiple step reactions (Sánchez-Jiménez et al., 2013).

54 Isoconversional methods can be divided into differential and integral methods (Wang et al., 2017). The

55 Friedman (FR) differential isoconversional model is among the most widely used (Burnham and Dinh,

56 2007). It has shown adequacy and high matching to experimental data in the pyrolysis of corn stalk (Cai et

al., 2018), and has been used to describe the kinetics of Miscanthus grass pyrolysis (Cortés and

58 Bridgwater, 2015). Kissinger-Akahira-Sunose (KAS) and Ozawa-Flynn-Wall (OFW) integral

59 isoconversional models have been applied in the pyrolysis of different grass types, such as Para grass (Al

60 Ayed et al., 2016) and Camel grass (Mehmood et al., 2017).

61 Product yields by pyrolysis of lignocellulosic biomass can be improved by pretreatment, such as by 62 the application of heat, chemicals or different pretreatment times (Wang et al., 2018) or by integration of 63 aerobic and anaerobic digestion (Juchelková et al., 2015). Pyrolysis is an attractive option for the 64 treatment of digestate due to its seasonal applicability as a fertilizer (Losak et al., 2014), and in cases when digestates contain heavy metals, pathogens and other toxic compounds (Wišniewski et al., 2015).

- 66 Pyrolysis of digestate is an interesting option because of the benefits it confers (Li et al., 2017) such as
- 67 reduction of weight and volume of digestate and elimination of pathogens and odour (Nansubuga et al.,

68 2015). As biochar from digestate can effectively be used for various applications, a combined anaerobic

- 69 digestion pyrolysis process might be beneficial because of the low economic value of digestate (Egieya
- ro et al., 2019); thus, subsequent pyrolysis of digestate offers an opportunity to improve the profitability of
- 71 biogas production processes (Egieya et al., 2018).

Digestate has recently attracted significant attention as a potential feedstock for pyrolysis (Wei et al., 2018). Various kinetic studies have been performed on the combined anaerobic digestion - pyrolysis process for biomass/waste material with its digestate, for corn stover and its digestate (Zhang et al., 2017), for food waste and its digestate (Opatokun et al., 2015), and for different organic solid wastes (sewage sludge, food waste, vinasse and cow manure) and their digestates (Li et al., 2017).

77 This paper presents the continuation of the previous experimental study on anaerobic digestion of 78 two types of roadside residue grass, residue grass from the uncultivated land (next to minor road) and 79 from the highway verge (Bedoić et al., 2019). This research contains two novel scientific contributions, 80 such as the study on pyrolysis of residue roadside grass and its digestate and the study on determination of 81 degraded organic matter during anaerobic process based on the analysis of thermogravimetric curves. 82 Estimation of the organic matter degradation is studied in relation to biochemical biogas potential of 83 grasses, which was determined previously (Bedoić et al., 2019). Two isoconversional models, differential 84 FR model and integral KAS model are used to quantify the impacts of anaerobic digestion of two types of 85 roadside residue grass on parameters of pyrolysis kinetics. Verification of the applied models based on the 86 experimental data and estimated kinetic parameters is finally conducted with the aim to reconstruct the 87 kinetic behaviour of studied feedstocks in the pyrolysis. The study determines the share of compounds in 88 two types of roadside grass and in its digestates without using any chemical solvents and performs the 89 research on the energy recovery of residue grass.

90 2. Materials and methods

In this section, we present an overview of the methods applied, along with a brief description of TGA; two further linear isoconversional models used in the study are introduced. Our research is linked with a previous investigation on anaerobic digestion of different types of residue grass (Bedoić et al., 2019). Regarding residue grass sampling, preparation and characterisation, and laboratory results from anaerobic digestion, readers are referred to Bedoić et al. (2019). 96

2.1. Substrates used in the study

We focused on the use of two roadside grass types. The first was collected on the verge next to a minor road (RG-MR), while the second was collected on the verge next to a highway (RG-H). In our previous study (Bedoić et al., 2019) they were marked as RG1 (now RG-MR) and RG3 (now RG-H). Both grasslands are located near Zagreb, the capital city of Croatia, on locations not suitable for food and/or feed purposes. Nine samples were collected on each of the grasslands examined, which were then stored in plastic bags in a vacuum in a freezer at -15° C (Bedoić et al., 2019).

103 The proximate and ultimate analysis and analysis of heavy metals presence of residue grasses have 104 been performed. The results, including results from field measurements, are presented in Bedoić et al. 105 (2019). Both types of grass (see RG1 and RG3 in Table 1 by Bedoić et al., 2019) showed similar chemical 106 composition; RG-H exhibited higher yield on both a fresh and dry basis, longer stem length, lower 107 moisture and ash contents, higher heating values and higher carbon content, compared to RG-MR. 108 Significant differences were obtained in terms of heavy metal concentrations, where RG-H showed much 109 higher values than RG-MR. On dry basis, RG-H sample contains 93.60 mg/kg of iron, 11.20 mg/kg of 110 zinc, 8.57 mg/kg of manganese and 4.55 mg/kg of copper. On the other side, RG-MR sample contains on 111 dry basis 54.40 mg/kg of iron, 5.86 mg/kg of zinc, 7.64 mg/kg of manganese and 3.72 mg/kg of copper. 112 Other elements like lead, cadmium, mercury and nickel have shown lower concentrations, below 1 mg/kg 113 of dry grass. As expected, higher concentrations of metals were detected for the grass collected in the 114 intense traffic area (highway verge). 115 For this study, raw grass samples were reduced into smaller pieces of approx. 3–6 cm in length and

115 For this study, raw grass samples were reduced into smaller pieces of approx. 5–6 cm in length and 116 were dried in a laboratory oven at 105°C until constant weight before use.

117 Besides the two types of roadside residue grass, their digestates obtained by mono-digestion were 118 used as substrates for the analysis. For anaerobic digestion, substrates were chopped into smaller pieces of 119 approx. 3–6 cm in length. They were placed in 250 mL batch reactors in triplicate together with inoculum, 120 where the ratio between inoculum and grass for anaerobic digestion was 1:1 on a dry basis. In total, 9 g of 121 total solids (TS) were added to each reactor, and the dry matter in each reactor was 6%. Anaerobic 122 digestion was performed for 42 days at 39°C. During the process of anaerobic mono-digestion no 123 inhibition could be observed, despite relatively high concentration of heavy metals in the studied grass 124 samples.

125 The digestates, RGD-MR - digestate of roadside grass collected on the verge next to the minor road 126 (marked as MRG1 in Bedoić et al., 2019), and RGD-H – digestate of roadside grass collected on the verge 127 next to the highway (marked as MRG3 in Bedoić et al., 2019) were used for this investigation. Before 128 performing TGA analysis, digestates were dried in a laboratory oven at 105°C until constant weight. More details regarding the residue grass substrates and anaerobic digestion process can be found inBedoić et al. (2019).

131

132 **2.2. Thermogravimetric analysis**

Thermogravimetric measurements of dried samples (RG-MR, RG-H, RGD-MR and RGD-H) were conducted using a TA Instruments Q500 at the heating rates β_i of 2.5, 5, 10, 15 and 20°C/min at a temperature range from 35 to 800°C under a steady flow of nitrogen (100 mL/min) to maintain an inert atmosphere. Samples weighing approximately 10 mg were used for the analysis. Three replicates were run for each sample, and the average value is reported. TGA data (mass weights and derivative mass weights) were recorded with respect to temperature and time for the five heating rates considered.

140 **2.3.** Analysis of hemicellulose and cellulose degradation

141 To calculate the degradation of hemicellulose (*hc*) and cellulose (*c*) in residue grass samples – 142 $X_{RG}(hc+c)$ (%) based on the analysis of thermogravimetric curves of grass and digestate, the following 143 relation was used:

$$X_{\rm RG}(hc+c) = \frac{\overline{x}_{\rm RG}(hc+c) - \overline{x}_{\rm RGD}(hc+c) \cdot \frac{\overline{x}_{\rm RG}(l)}{\overline{x}_{\rm RGD}(l)}}{\overline{x}_{\rm RG}(hc+c)} \times 100$$
(1)

where $\bar{x}_{RG}(hc+c)$ is the average share of hemicellulose and cellulose in residue grass sample (before anaerobic digestion), $\bar{x}_{RGD}(hc+c)$ is the average share of hemicellulose and cellulose in digestate sample (after anaerobic digestion), $\bar{x}_{RG}(l)$ is the average share of lignin in residue grass sample (before anaerobic digestion) and $\bar{x}_{RGD}(l)$ is the average share of lignin in digestate sample (after anaerobic digestion). The term "average share of components" is related to the arithmetic mean of a component's share determined at the selected heating rates. Factor $\frac{\bar{x}_{RG}(l)}{\bar{x}_{RGD}(l)}$ is introduced in the calculation since the relative share of components during anaerobic digestion changes.

151

152 **2.4. Analysis of kinetic parameters**

153 Two linear isoconversional models (Li et al., 2017) were used for the determination of kinetic 154 parameters of roadside grass and roadside grass digestate pyrolysis, as shown in Table 1. In Table 1 β_i is 155 the heating rate (°C/min), α is the degree of conversion (/ or %), T is the temperature as a general

parameter of process (°C or K), $(d\alpha/dT)_{\alpha,i}$ is the conversion derivative per temperature at the given degree

157 of conversion and heating rate, $A_{\alpha}f(\alpha)$ is a modified pre-exponential factor in the Friedmann

isoconversional model (1/s), $A_{\alpha}/g(\alpha)$ is the modified pre-exponential factor in the Kissinger-Akahira-

159 Sunose isoconversional model (1/s), E_{α} is the activation energy (J/mol), $T_{\alpha,i}$ is the temperature at the given

160 degree of conversion and heating rate required for model application (K), and R is the universal gas

161 constant (8.314 J/(mol·K)).

162

163 Table 1: Linear isoconversional kinetic models applied on the pyrolysis of roadside grass and roadside164 grass digestate

165

166 To conduct a kinetic analysis using the models in Table 1, it is necessary to determine the degree of 167 conversion at a certain temperature $\alpha(T)$, as:

$$\alpha(T) = \frac{m_0 - m(T)}{m_0 - m_f}$$
(2)

where m_0 is the mass at temperature T_0 , m(T) is the mass at temperature T, and m_f is the mass at the final temperature T_f . Since biomass sample contains retained water and could also contain light volatile compounds, the first stage is a dehydration stage. Pyrolysis occurs in the second and third stages, which are called active and passive pyrolysis. Both pyrolysis stages correspond to the decomposition of cellulose, hemicellulose and lignin. It has been reported that the dehydration stage ends at about 150°C, and pyrolysis of lignocellulosic biomass starts (Chen et al., 2013). When applying the Friedman isoconversional model in the kinetic study of pyrolysis, it is necessary

to determine the derivative conversion curve $(d\alpha/dT)_{\alpha,i}$. Since the curve has been reported to have many fluctuations, it is recommended to apply some smoothing tool to reduce the impact of the noisy data (Vyazovkin et al., 2011). In this study, the Moving Average of data in Excel was applied (Hogarth, 2014) to smooth the experimental derivative conversion curves.

After the degree of conversion has been determined, the calculations of parameters based on the thermogravimetric measurements follows. For the Friedman isoconversional model, at the given α , E_{α} and $\ln[A_{\alpha}f(\alpha)]$ are obtained from the slope and intercept of the plot of $\ln[\beta_i(d\alpha/dT)_{\alpha,i}]$ versus $(-1/RT_{\alpha,i})$. For the Kissinger-Akahira-Sunose isoconversional model, at the given α , E_{α} and $\ln[A_{\alpha}/g(\alpha)]$ are obtained from the slope and intercept of the plot of $\ln[\beta_i/T^2_{\alpha,i}]$ versus $(-1/RT_{\alpha,i})$.

184 To represent the deviations of activation energy and modified pre-exponential factor (based on 185 linear regression) at the given degree of conversion, the confidence interval has been used (Cai et al.,

186	2018). The specific level of confidence was set at 95% to present a range of values of activation energy
187	and modified pre-exponential factor within the selected probability (Bartocci et al., 2019).
188	To reconstruct the kinetic behaviour of pyrolysis of the selected feedstocks, the average values of
189	kinetic parameters at the given degree of conversion are used.
190	3. Results and discussion
191	In this section, the results of our experiments of biomass pyrolysis and modelling of pyrolysis
192	kinetics are presented.
193	
194	3.1. Thermogravimetric analysis
195	The results of thermogravimetric analysis of roadside grass (RG) and roadside grass digestate
196	(RGD) in terms of thermogravimetric (TG) and derivative TG (DTG) curves are presented in Figure 1 for
197	five different heating rates (β_i of 2.5, 5, 10, 15 and 20°C/min), where the temperature range is from 35 to
198	800°C. TG shows the loss of weight during heating, while DTG shows the first derivative of TG which
199	indicates the main devolatilization stages more clearly (Ceylan and Kazan, 2015). TG curves for analysed
200	samples (RG-MR, RG-H, RGD-MR and RGD-H) show steady or decreasing trends with increased
201	temperature, while the changes in TG curves (weight loss) are shown as peaks in DTG curves.
202	
203	Figure 1: TG-DTG curves of RG and RGD samples at different heating rates: a) 2.5 °C/min, b) 5 °C/min,

c) 10 °C/min, d) 15 °C/min, e) 20 °C/min

206 All the samples underwent three main stages of weight loss, which indicate the processes of 207 dehydration, active and passive pyrolysis (Slopiecka et al., 2012). The stages are the drying, 208 devolatilization and char formation stages (Chandrasekaran et al., 2017). In the dehydration stage, 209 evaporation of water and light volatile compounds occurs; in the active pyrolysis stage, the degradation of 210 hemicellulose and cellulose takes place, and in the final stage, decomposition of lignin occurs 211 (Chandrasekaran et al., 2017). Hemicellulose and cellulose degrade at a similar temperature range 212 (simultaneously) and thus only one peak is typically obtained in the DTG curve (Parthasarathy and 213 Narayanan, 2014). The last stage typically shows slow continued loss of weight, as solid residue is slowly 214 decomposed (Peng et al., 2001). The final residue at 800°C consists of biochar and ash (Peng et al., 2001). 215 In Figure 1 it can be seen that RG-H shows the highest peak of the DTG curve at the highest 216 temperature at each of the applied heating rates, and exhibits a more intense peak shoulder compared to

204

205

217 RG-MR. Unlike the RG samples, peak shoulder in the DTG curves in the case of RGDs is not clearly

- visible. We assume that this is because both cellulose and hemicellulose from RG are partly degraded
- 219 under anaerobic conditions, which impact the DTG curves of RGD by fading the peak shoulder. Similar
- 220 observations have been reported in the analysis of food waste and its digestate pyrolysis (Opatokun et al.,

221 2015).

According to the TG curves, all the RGDs show higher amounts of residues or lower weight loss during pyrolysis due to lower cellulose and hemicellulose content in the samples. In terms of grass samples, RG-MR shows a slightly higher yield of the final residue compared to RG-H. This result can be explained through the higher share of ash and carbon in the RG-MR sample (10.4% of ash and 47.1% of carbon, on dry basis) than in the RG-H sample (8.4% of ash and 46.2% of carbon, on dry basis) (Bedoić et al., 2019).

228 More detailed information regarding the mass loss intervals and characteristic temperature zones 229 (Ye et al., 2010) of all samples is shown in Tables 2 and 3. Table 2 shows five characteristic temperature 230 zones during different stages of decomposition of RG and RGD samples at five different heating rates. T1 231 in Table 2 represents the end of the dehydration stage or the start of the biomass pyrolysis process. The 232 active pyrolysis stage occurs at temperatures between T1 and T5. This stage (stage II) can be divided into zones I and II, where zone I occurs at temperatures T1 - T3, and zone II at temperatures T3 - T5, with 233 234 maximum weight loss at T2 and T4. For more details regarding characteristic temperature zones, see (Ye 235 et al., 2010). As mentioned previously, the upper limit of the temperature range, 800°C, is applied.

236 237

 Table 2: Characteristic temperature zones during different stages of decomposition of RG and RGD samples

239 240

241

238

Table 3: Weight loss (in wt.%) during different stages of decomposition of RG and RGD samples

Results show that the dehydration stage occurs from the starting temperature to about 136 to 191°C for RG samples, and to about 137 to 189°C for RGD samples. The active pyrolysis stage is observed to be in the following temperature ranges:

245 • RG-MR: start from ca. 136°C (2.5 °C/min) to ca. 191°C (20°C/min), end from ca. 343°C 246 (2.5°C/min) to ca. 396°C (20°C/min),

• RG-H: start from ca. 139°C (2.5°C/min) to ca. 191°C (20°C/min), end from ca. 356°C

248 (2.5°C/min) until ca. 418°C (20°C/min),

249 • RGD-H: start from ca. 137°C (2.5°C/min) to ca. 187.5°C (20 °C/min), end from ca. 332°C
250 (2.5°C/min) until ca. 394°C (20°C/min),

251

• RGD-H: start from ca. 141°C (2.5°C/min) to ca. 189°C (20°C/min), end from ca. 364°C

252 (2.5°C/min) until ca. 415°C (20°C/min),

253 Table 3 shows the weight loss during different stages of decomposition for all the analysed samples 254 for the five heating rates considered. From Table 3 it can be seen that the first dehydration stage shows 255 slightly higher weight loss for RGD samples (ca. 6 to 8%) than for RG samples (ca. 4 to 5.5%). RG-H 256 samples on average contain slightly more cellulose and hemicellulose, $\bar{x}_{RG-H}(hc+c)=60\%$, compared to 257 RG-MR samples, $\bar{x}_{RG-MR}(hc+c)=56\%$. Since temperatures T2 and T3 could not be determined for the RGD 258 samples due to the shoulder fading in DTG curves, the share of cellulose and hemicellulose is calculated 259 by subtracting the share of moisture, lignin and final residue from the total amount (100 wt.%). Therefore, 260 the amount of hemicellulose and cellulose in digestate samples is estimated on average, \bar{x}_{RGD}

261 $_{MR}(hc+c)=37\%$ and $\bar{x}_{RGD-H}(hc+c)=42\%$.

262 The degradation of lignin at the observed heating rates started at ca. 340 to 400°C in the RG-MR sample, at ca. 360 to 420°C for the RG-H sample, at ca. 330 to 390°C for the RGD-MR sample and at ca. 263 360 to 415°C for the RGD-H sample. The mass loss caused by lignin degradation and charring is as 264 265 follows: $\bar{x}_{\text{RGD-MR}}(l)=17\%$, $\bar{x}_{\text{RG-MR}}(l)=13\%$, $\bar{x}_{\text{RGD-H}}(l)=15\%$, and $\bar{x}_{\text{RG-H}}(l)=12\%$. The differences in mass loss 266 during the lignin degradation and biochar formation stage between RGD samples and raw RG samples are 267 3 and 4%, on average. Since the relative share of components during anaerobic digestion changes, this 268 could be the reason for the measured deviations. However, estimation of the amount of lignin in samples 269 should be taken with caution, since it was obtained by using experimental data and theoretical background 270 related to pyrolysis of lignocellulosic biomass (Carrier et al., 2016). It has been reported that partial 271 degradation of lignin under inert atmosphere starts at 200°C, while at 400°C it starts to be intensified 272 (Carrier et al., 2011).

When Eq. (1) is applied, the results show that during monodigestion of RG-MR, ca. 50% of cellulose and hemicellulose was converted to biogas ($X_{\text{RG-MR}}(hc+c)=49.5\%$), and in the case of RG-H, degradation of cellulose and hemicellulose is estimated at $X_{\text{RG-H}}(hc+c)=44.0\%$). Degradation is similar for both grass types, which is also supported by the fact that for both, similar biochemical biogas potential values (BGP) were obtained, 0.436 Nm³/kgTS for RG-MR and 0.413 Nm³/kgTS for RG-H (Bedoić et al., 2019).

The results show that mono-digestion has been incomplete (49.5 and 44 % conversion of cellulose and hemicellulose). To further increase the degradability of biomass and enhance biogas production, codigestion and addition of additives such as bio-based carbon materials (Yun et al., 2018) and accelerants such as for example urea, plant ash (Zhang et al., 2018) or steel slag (Han et al., 2019) to substrates have been recognised to be more efficient than mono-digestion (Wang et al., 2019). Similar conclusions have been obtained by thermogravimetric analysis of digested residue from aloe peel waste and dairy manure(Huang et al., 2016).

At 800°C, the average final residue yields for RG-MR and RG-H are ca. 25% and 23%. However, RGD samples have shown a higher yield of final residue at 800°C; for RGD-MR, the yield was ca. 38%, and for RGD-H, ca. 37%. Both RGD-MR and RGD-H have shown much higher yields of the final residues at 800°C than the RG feedstocks. Similar conclusions were obtained when using food waste as feedstock (Opatokun et al., 2015).

This study has shown that significant quantities of final residue (mainly biochar) are obtained from pyrolysis of roadside grass and its digestate. Biochar could have various applications, such as it could be used as an additive material for improving stability of anaerobic digestion, as an approach to carbon sequestration, in animal husbandry, as a soil conditioner, in the building sector, in treatment of drinking and waste waters and in many other applications (Schmidt, 2012). Combined anaerobic digestion – pyrolysis process from roadside grass might also improve the profitability of biogas production processes (Egieva et al., 2018).

298

3.2. Kinetic analysis

300 Thermogravimetry and isoconversional models can provide an estimation of kinetic data (activation 301 energy and pre-exponential factor) from reaction parameters such as temperature and heating rate without 302 estimation of reaction mechanisms (Damartzis et al., 2011). The activation energy and modified pre-303 exponential factors were obtained using FR and KAS methods. Their distribution for RG and RGD 304 samples has been determined based on the performed thermogravimetric analysis data for conversions 305 between 20 and 70% in step sizes of 5%. Degrees of conversion lower than 20% and higher than 70% are 306 not shown because of significant fluctuations observed (especially for digestate samples), which were 307 probably associated with the thermal behaviour of lignin (Carrier et al., 2016). In addition, verification of 308 the applied models was performed, and average values of kinetic parameters obtained by this study were 309 used to verify models with the experimental data.

310

3.2.1. Friedman (FR) model

The values of activation energy (E_{α}) and modified pre-exponential factor in logarithmic expression (ln[$A_{\alpha}f(\alpha)$]) for pyrolysis of RG and RGD samples were obtained using the FR isoconversional model, as shown in Figure 2. The error bars in Figure 2 represent confidence intervals with a confidence level of 95%.

315

1	~ 1
316	Figure 2: Distribution of E_{α} and $\ln[A_q f(\alpha)]$ per the degree of conversion by means of FR model
317	
318	Estimated values of E_{α} in the studied range of conversions vary between 160 and 600 kJ/mol for
319	RG-MR samples, and between 170 and 380 kJ/mol for RG-H samples. Both RG samples show a slight
320	increase in the values of E_{α} from $\alpha = 0.20$ to 0.30; between $\alpha = 0.30$ to 0.50, a stagnation/slight decline of
321	E_{α} is shown, and after $\alpha = 0.50$, a significant increase in the E_{α} can be observed. Such a trend in the
322	distribution of E_{α} using the FR model was also reported for corn stalk pyrolysis (Cai et al., 2018) and for
323	miscanthus pyrolysis (Cortés and Bridgwater, 2015).
324	On the other hand, RGD samples have shown much lower values of E_{α} in the considered ranges of
325	conversions; for RGD-MR it is between 20 and 170 kJ/mol, while for RGD-H it is between 10 and 170
326	kJ/mol. RGD samples show the highest E_{α} at the lowest value of α , and with an increase in the degree of
327	conversion, E_{α} continuously declines in the case of RGD-MR, while RGD-H declines up to $\alpha = 0.45$, and
328	then stagnation appears.
329	Similar trends as for E_{α} are observed for the change of $\ln[A_{\alpha}f(\alpha)]$ with the degree of conversion. The
330	highest value of $\ln[A_{\alpha}f(\alpha)]$ is for RG-MR, about 110 s ⁻¹ , while the highest value for RG-H is around 60 s ⁻¹ .
331	RGD samples show negative values of $\ln[A_{\alpha}f(\alpha)]$, with the lowest value around -8 s ⁻¹ .
332	To reconstruct the kinetic process using the FR model, Eq. (3) is used in the non-logarithmic form.
333	The results of the verification process are presented in Figure 3.
334	
335	Figure 3: Experimental and FR kinetic model based on E_{α} and $\ln[A_{\alpha}f(\alpha)]$ data for grass and digestate
336	pyrolysis at 2.5, 5, 10, 15 and 20 °C/min
337	
338	At all heating rates, the FR model shows high-level matching with the experimental data for RG
339	samples. Higher fluctuations of the model compared to the experimental data are shown for RGD samples.
340	The peaks of the curves for RG samples move to higher temperatures with an increase in heating rate.
341	That observation is not seen for RGD samples. Furthermore, RGD samples show a wider range of
342	temperatures in terms of the $\beta_i(d\alpha/dT)$ distribution. Since the weight loss in the active pyrolysis stage
343	(stage II) is significantly lower for RGD samples, and the residue yield greater than for RG samples, this
344	observation is supported by the analysis of TG curves. The kinetic parameters obtained with the FR model
345	show a better fit for RG samples than for RGD samples. At lower heating rates of 2.5 and 5°C/min, RGD
346	samples show a good model fit to the experimental data.

347 3.2.2. Kissinger-Akahira-Sunose (KAS) model

348 Using the KAS isoconversional model, the values of activation energy (E_{α}) and modified pre-349 exponential factor in logarithmic expression $(\ln[A_{\alpha}/g(\alpha)])$ for pyrolysis of RG and RGD samples are 350 obtained, as shown in Figure 4**Error! Reference source not found.**

351

352

353

Figure 4: Distribution of E_{α} and $\ln[A_{\alpha}/g(\alpha)]$ with the degree of conversion by means of KAS model

354 E_{α} estimated by the KAS model for RG samples in the studied range of conversions vary between 355 150 and 430 kJ/mol for RG-MR, and between 160 and 260 kJ/mol for RG-H samples. Similar results were 356 obtained for the pyrolysis of Para grass (between 180 and 230 kJ/mol, (Al Ayed et al., 2016)) and Camel 357 grass with the KAS model (between 150 and 190 kJ/mol, (Mehmood et al., 2017)). The results obtained in 358 this study and by analyses of specific grass types show a narrower range of activation energies for specific 359 grass types than for unclassified species of grass. On the other hand, RGD samples again show lower 360 values of E_{α} compared to RG samples; for RGD-MR the range is between 30 and 170 kJ/mol, while for 361 RGD-H it is between 20 and 175 kJ/mol. Again, RGD samples show the highest E_{α} at the lowest value of 362 α . KAS modeling shows that with an increase of the degree of conversion, E_{α} continuously declines in the 363 case of both RGD samples. 364 Similar trends with the degree of conversion as for E_{α} are obtained for $\ln[A_{\alpha}/g(\alpha)]$ for both RG and 365 RGD samples. The highest value of $\ln[A_{\alpha}/g(\alpha)]$ is obtained for RG-MR, about 80 s⁻¹, while the highest

value for RG-H is about 45 s⁻¹. It should be noted that modified pre-exponential factors obtained with FR and KAS models cannot be compared directly, since the expressions of functions are slightly different. RGD samples again show both positive and negative values of $\ln[A_{\alpha}/g(\alpha)]$, where the lowest value is about -5 s⁻¹.

- To reconstruct the kinetic process with the KAS model, Eq. (4) is used in the non-logarithmic form.
 The results of the verification process are shown in Figure 5.
- 372

373 Figure 5: Experimental and KAS kinetic model based on E_{α} and $\ln[A_{\alpha}/g(\alpha)]$ for grass and digestate 374 pyrolysis at 2.5, 5, 10, 15 and 20 °C/min

375

The KAS model shows matching with the experimental data for the middle temperature range of 450 to 700 K in the case of RG samples, and between 500 and 1,050 K for RGD samples. At lower temperatures, the KAS model deviates significantly from the experimental data. Again, more intense fluctuations of the model compared to the experimental data are shown for RGD samples. For all the analysed samples, the highest match of KAS model to experimental data is at the lowest heating rate

- 381 (2.5°C/min). Slowly heating the samples leads to a better and more effective heat transfer to the inner
- 382 layers of biomass (Mani et al., 2010). Therefore, the model results match the experimental data best at
- 383 lower heating rates. Kinetic parameters obtained with the KAS model are more effective in the case of RG
- 384 samples than in the case of its digestate.

385 4. Conclusions

- The analysis of TG and DTG curves of selected feedstocks shows that estimated amount of degraded cellulose and hemicellulose in roadside grass during the AD process is around 44 to 50%. Roadside grass digestate has shown a greater yield of final residues (ca. 38%) than roadside grass samples (ca. 24%). The combined process, anaerobic digestion of roadside grass and pyrolysis of its digestate, contributes to the production of green bioenergy in the form of heat and electricity, while reducing energy requirements (activation energy and pre-exponential factor) for pyrolysis.
- 392

393 Acknowledgements

The authors acknowledge financial support from the European Union's Horizon 2020 research and innovation programme under grant agreement No: 690142-2 (AgroCycle project), from the Slovenian Research Agency (core research funding No. P2 0412) and from the Slovenia-Croatia bilateral project Integration of renewable energy within energy systems (INTEGRES).

398

399 Symbols

Conversion derivative per temperature at the given degree of conversion and heating
rate
Initial (mass or temperature)
Modified pre-exponential factor in KAS isoconversional model (s ⁻¹)
Modified pre-exponential factor in FR isoconversional model (s ⁻¹)
Final (mass or temperature)
mass (kg)
Universal gas constant, 8.314 J/(mol·K)
Temperature, general parameter of process (°C or K)
Temperature at the given degree of conversion and heating rate required for model
application (K)
degradation of hemicellulose and cellulose during anaerobic digestion of roadside grass

	(/ or %)
\bar{x}	average share of a component in a biomass sample (/ or %)
α	Degree of conversion (/or %)
β_i	Heating rate (°C/min)

400 **References**

- Akhtar, A., Krepl, V., Ivanova, T., 2018. A Combined Overview of Combustion, Pyrolysis, and
 Gasification of Biomass. Energy and Fuels 32, 7294–7318.
 https://doi.org/10.1021/acs.energyfuels.8b01678
- Al Ayed, O.S., Rashid, U., Ye, G., Luo, H., Mehmood, M.A., Ibrahim, M., Arbi Nehdi, I., Qadir,
 G., Ahmad, M.S., 2016. Kinetic analyses and pyrolytic behavior of Para grass (Urochloa mutica)
 for its bioenergy potential. Bioresour. Technol. 224, 708–713.
 https://doi.org/10.1016/j.biortech.2016.10.090
- Bartocci, P., Tschentscher, R., Stensrød, R.E., Barbanera, M., Fantozzi, F., 2019. Kinetic Analysis
 of Digestate Slow Pyrolysis with the Application of the Master-Plots Method and Independent
 Parallel Reactions Scheme. Molecules 24, 1657. https://doi.org/10.3390/molecules24091657
- 4. Bedoić, R., Čuček, L., Ćosić, B., Krajnc, D., Smoljanić, G., Kravanja, Z., Ljubas, D., Pukšec, T.,
 Duić, N., 2019. Green biomass to biogas A study on anaerobic digestion of residue grass. J.
 Clean. Prod. 213, 700–709. https://doi.org/10.1016/j.jclepro.2018.12.224
- 414 5. Burnham, A.K., Dinh, L.N., 2007. A comparison of isoconversional and model-fitting approaches
 415 to kinetic parameter estimation and application predictions. J. Therm. Anal. Calorim. 89, 479–490.
 416 https://doi.org/10.1007/s10973-006-8486-1
- Cai, J., Xu, D., Dong, Z., Yu, X., Yang, Y., Banks, S.W., Bridgwater, A. V., 2018. Processing
 thermogravimetric analysis data for isoconversional kinetic analysis of lignocellulosic biomass
 pyrolysis: Case study of corn stalk. Renew. Sustain. Energy Rev. 82, 2705–2715.
 https://doi.org/10.1016/j.rser.2017.09.113
- 421 7. Campuzano, F., Brown, R.C., Martínez, J.D., 2019. Auger reactors for pyrolysis of biomass and
 422 wastes. Renew. Sustain. Energy Rev. 372–409. https://doi.org/10.1016/j.rser.2018.12.014
- 423 8. Carrier, M., Auret, L., Bridgwater, A., Knoetze, J.H., 2016. Using Apparent Activation Energy as a
 424 Reactivity Criterion for Biomass Pyrolysis. Energy and Fuels 30, 7834–7841.
 425 https://doi.org/10.1021/acs.energyfuels.6b00794
- 426 9. Carrier, M., Loppinet-Serani, A., Denux, D., Lasnier, J.M., Ham-Pichavant, F., Cansell, F.,
 427 Aymonier, C., 2011. Thermogravimetric analysis as a new method to determine the lignocellulosic

- 428 composition of biomass. Biomass and Bioenergy 35, 298–307.
 429 https://doi.org/10.1016/j.biombioe.2010.08.067
- 430 10. Ceylan, S., Kazan, D., 2015. Pyrolysis kinetics and thermal characteristics of microalgae
 431 Nannochloropsis oculata and Tetraselmis sp. Bioresour. Technol. 187, 1–5.
 432 https://doi.org/10.1016/j.biortech.2015.03.081
- 433 11. Chandrasekaran, A., Ramachandran, S., Subbiah, S., 2017. Determination of kinetic parameters in
 434 the pyrolysis operation and thermal behavior of Prosopis juliflora using thermogravimetric
 435 analysis. Bioresour. Technol. 233, 413–422. https://doi.org/10.1016/j.biortech.2017.02.119
- 436 12. Chen, D., Zheng, Y., Zhu, X., 2013. In-depth investigation on the pyrolysis kinetics of raw
 437 biomass. Part I: Kinetic analysis for the drying and devolatilization stages. Bioresour. Technol.
 438 131, 40–46. https://doi.org/10.1016/j.biortech.2012.12.136
- 439 13. Cortés, A.M., Bridgwater, A. V., 2015. Kinetic study of the pyrolysis of miscanthus and its acid
 440 hydrolysis residue by thermogravimetric analysis. Fuel Process. Technol. 138, 184–193.
 441 https://doi.org/10.1016/j.fuproc.2015.05.013
- 442 14. Damartzis, T., Vamvuka, D., Sfakiotakis, S., Zabaniotou, A., 2011. Thermal degradation studies
 443 and kinetic modeling of cardoon (Cynara cardunculus) pyrolysis using thermogravimetric analysis
 444 (TGA). Bioresour. Technol. 102, 6230–6238. https://doi.org/10.1016/j.biortech.2011.02.060
- 445 15. Dhyani, V., Bhaskar, T., 2018. A comprehensive review on the pyrolysis of lignocellulosic
 446 biomass. Renew. Energy 129, 695–716. https://doi.org/10.1016/j.renene.2017.04.035
- 447 16. Egieya, J.M., Čuček, L., Zirngast, K., Isafiade, A.J., Pahor, B., Kravanja, Z., 2019. Synthesis of
 448 biogas supply networks using various biomass and manure types. Comput. Chem. Eng. 122, 129–
 449 151. https://doi.org/10.1016/j.compchemeng.2018.06.022
- 450 17. Egieya, J.M., Čuček, L., Zirngast, K., Isafiade, A.J., Pahor, B., Kravanja, Z., 2018. Biogas supply
 451 chain optimization considering different multi-period scenarios. Chem. Eng. Trans. 70, 985–990.
 452 https://doi.org/10.3303/CET1870165
- 453 18. Han, F., Yun, S., Zhang, C., Xu, H., Wang, Z., 2019. Steel slag as accelerant in anaerobic digestion
 454 for nonhazardous treatment and digestate fertilizer utilization. Bioresour. Technol. 331–338.
 455 https://doi.org/10.1016/j.biortech.2019.03.029
- 456 19. Hogarth, M., 2014. Microsoft Excel techniques. Data Clean-Up Manag. 43–81.
 457 https://doi.org/10.1016/b978-1-84334-672-2.50005-6
- 458 20. Huang, X., Yun, S., Zhu, J., Du, T., Zhang, C., Li, X., 2016. Mesophilic anaerobic co-digestion of
 459 aloe peel waste with dairy manure in the batch digester: Focusing on mixing ratios and digestate
 460 stability. Bioresour. Technol. 218, 62–68. https://doi.org/10.1016/j.biortech.2016.06.070

- 461 21. Jain, A.A., Mehra, A., Ranade, V. V., 2016. Processing of TGA data: Analysis of isoconversional
 462 and model fitting methods. Fuel 165, 490–498. https://doi.org/10.1016/j.fuel.2015.10.042
- 463 22. Juchelková, D., Corsaro, A., Hlavsová, A., Raclavská, H., 2015. Effect of composting on the
 464 production of syngas during pyrolysis of perennial grasses. Fuel 154, 380–390.
 465 https://doi.org/10.1016/j.fuel.2015.03.061
- 466 23. Li, X., Mei, Q., Dai, X., Ding, G., 2017. Effect of anaerobic digestion on sequential pyrolysis
 467 kinetics of organic solid wastes using thermogravimetric analysis and distributed activation energy
 468 model. Bioresour. Technol. 227, 297–307. https://doi.org/10.1016/j.biortech.2016.12.057
- 469 24. Losak, T., Hlusek, J., Zatloukalova, A., Musilova, L., Vitezova, M., Skarpa, P., Zlamalova, T.,
 470 Fryc, J., Vitez, T., Marecek, J., Martensson, A., 2014. Digestate from Biogas Plants is an Attractive
 471 Alternative to Mineral Fertilisation of Kohlrabi. J. Sustain. Dev. Energy, Water Environ. Syst. 2,
 472 309–318. https://doi.org/10.13044/j.sdewes.2014.02.0025
- 473 25. Malherbe, S., Cloete, T.E., 2002. Lignocellulose biodegradation: Fundamentals and applications.
 474 Rev. Environ. Sci. Biotechnol. 1, 105–114. https://doi.org/10.1023/A:1020858910646
- 475 26. Mani, T., Murugan, P., Abedi, J., Mahinpey, N., 2010. Pyrolysis of wheat straw in a 476 thermogravimetric analyzer: Effect of particle size and heating rate on devolatilization and 477 of 88. estimation global kinetics. Chem. Eng. Res. Des. 952-958. 478 https://doi.org/10.1016/j.cherd.2010.02.008
- 479 27. Mehmood, M.A., Ye, G., Luo, H., Liu, C., Malik, S., Afzal, I., Xu, J., Ahmad, M.S., 2017.
 480 Pyrolysis and kinetic analyses of Camel grass (Cymbopogon schoenanthus) for bioenergy.
 481 Bioresour. Technol. 228, 18–24. https://doi.org/10.1016/j.biortech.2016.12.096
- 482 28. Mikulcic, H., Jin, Q., Stančin, H., Wang, X., Li, S., Tan, H., Duić, N., 2019. Thermogravimetric 483 Analysis Investigation of Polyurethane Plastic Thermal Properties Under Different Atmospheric 484 Conditions. J. Sustain. Energy, Water Environ. Syst. 7. 355-367. Dev. 485 https://doi.org/10.13044/j.sdewes.d6.0254
- 486 29. Nansubuga, I., Banadda, N., Ronsse, F., Verstraete, W., Rabaey, K., 2015. Digestion of high rate
 487 activated sludge coupled to biochar formation for soil improvement in the tropics. Water Res. 81,
 488 216–222. https://doi.org/10.1016/j.watres.2015.05.047
- 489 30. Opatokun, S.A., Strezov, V., Kan, T., 2015. Product based evaluation of pyrolysis of food waste
 490 and its digestate. Energy 92, 349–354. https://doi.org/10.1016/j.energy.2015.02.098
- 491 31. Oyedun, A.O., Tee, C.Z., Hanson, S., Hui, C.W., 2014. Thermogravimetric analysis of the
 492 pyrolysis characteristics and kinetics of plastics and biomass blends. Fuel Process. Technol. 128,
 493 471–481. https://doi.org/10.1016/j.fuproc.2014.08.010

- 494 32. Parthasarathy, P., Narayanan, S.K., 2014. Determination of kinetic parameters of biomass samples
 495 using thermogravimetric analysis. Environ. Prog. Sustain. Energy 33, 256–266.
 496 https://doi.org/10.1002/ep.11763
- 497 33. Peng, W., Wu, Q., Tu, P., Zhao, N., 2001. Pyrolytic characteristics of microalgae as renewable
 498 energy source determined by thermogravimetric analysis. Bioresour. Technol. 80, 1–7.
 499 https://doi.org/10.1016/S0960-8524(01)00072-4
- 34. Ramajo-Escalera, B., Espina, A., García, J.R., Sosa-Arnao, J.H., Nebra, S.A., 2006. Model-free
 kinetics applied to sugarcane bagasse combustion. Thermochim. Acta 448, 111–116.
 https://doi.org/10.1016/j.tca.2006.07.001
- 503 35. Rodriguez, C., Alaswad, A., Benyounis, K.Y., Olabi, A.G., 2017. Pretreatment techniques used in
 504 biogas production from grass. Renew. Sustain. Energy Rev. 68, 1193–1204.
 505 https://doi.org/10.1016/j.rser.2016.02.022
- 36. Rostek, E., Biernat, K., 2013. Thermogravimetry as a Research Method in the Transformation
 Processes of Waste Rubber and Plastic Products for Energy Carriers (WtE and WtL Processes). J.
 Sustain. Dev. Energy, Water Environ. Syst. 1, 163–171.
 https://doi.org/10.13044/j.sdewes.2013.01.0012
- 510 37. Sánchez-Jiménez, P.E., Pérez-Maqueda, L.A., Perejón, A., Criado, J.M., 2013. Clarifications
 511 regarding the use of model-fitting methods of kinetic analysis for determining the activation energy
 512 from a single non-isothermal curve. Chem. Cent. J. 7. https://doi.org/10.1186/1752-153X-7-25
- 513 38. Schmidt, H.-P., 2012. The 55 uses of biochar. Ithaka J. 1, 286–289. www.ithaka-journal.net.
- 514 39. Senneca, O., 2007. Kinetics of pyrolysis, combustion and gasification of three biomass fuels. Fuel
 515 Process. Technol. 88, 87–97. https://doi.org/10.1016/j.fuproc.2006.09.002
- 40. Slopiecka, K., Bartocci, P., Fantozzi, F., 2012. Thermogravimetric analysis and kinetic study of
 poplar wood pyrolysis. Appl. Energy 97, 491–497. https://doi.org/10.1016/j.apenergy.2011.12.056
- 518 41. Stančin, H., Růžičková, J., Mikulčić, H., Raclavská, H., Kucbel, M., Wang, X., Duić, N., 2019.
 519 Experimental analysis of waste polyurethane from household appliances and its utilization
 520 possibilities. J. Environ. Manage. 243, 105–115. https://doi.org/10.1016/j.jenvman.2019.04.112
- 42. Vyazovkin, S., Burnham, A.K., Criado, J.M., Pérez-Maqueda, L.A., Popescu, C., Sbirrazzuoli, N.,
 2011. ICTAC Kinetics Committee recommendations for performing kinetic computations on
 thermal analysis data. Thermochim. Acta 520, 1–19. https://doi.org/10.1016/j.tca.2011.03.034
- 43. Wang, S., Dai, G., Yang, H., Luo, Z., 2017. Lignocellulosic biomass pyrolysis mechanism: A
 state-of-the-art review. Prog. Energy Combust. Sci. 62, 33–86.
 https://doi.org/10.1016/j.pecs.2017.05.004

17

- 44. Wang, X., Bai, S., Jin, Q., Li, S., Li, Yukun, Li, Yan, Tan, H., 2018. Soot formation during
 biomass pyrolysis: Effects of temperature, water-leaching, and gas-phase residence time. J. Anal.
 Appl. Pyrolysis 134, 484–494. https://doi.org/10.1016/j.jaap.2018.07.015
- 45. Wang, Z., Yun, S., Xu, H., Wang, C., Zhang, Y., Chen, J., Jia, B., 2019. Mesophilic anaerobic codigestion of acorn slag waste with dairy manure in a batch digester: Focusing on mixing ratios and
 bio-based carbon accelerants. Bioresour. Technol. https://doi.org/10.1016/j.biortech.2019.121394
- 46. Wei, Y., Hong, J., Ji, W., 2018. Thermal characterization and pyrolysis of digestate for phenol
 production. Fuel 232, 141–146. https://doi.org/10.1016/j.fuel.2018.05.134
- 47. Wišniewski, D., Gołaszewski, J., Białowiec, A., 2015. The pyrolysis and gasification of digestate
 from agricultural biogas plant. Arch. Environ. Prot. 41, 70–75. https://doi.org/10.1515/aep-20150032
- 48. Yun, S., Fang, W., Du, T., Hu, X., Huang, X., Li, X., Zhang, C., Lund, P.D., 2018. Use of biobased carbon materials for improving biogas yield and digestate stability. Energy 164, 898–909.
 https://doi.org/10.1016/j.energy.2018.09.067
- 49. Zhang, C., Yun, S., Li, X., Wang, Z., Xu, H., Du, T., 2018. Low-cost composited accelerants for
 anaerobic digestion of dairy manure: Focusing on methane yield, digestate utilization and energy
 evaluation. Bioresour. Technol. 263, 517–524. https://doi.org/10.1016/j.biortech.2018.05.042
- 54. 50. Zhang, D., Wang, F., Yi, W., Li, Z., Shen, X., Niu, W., 2017. Comparison study on pyrolysis
 characteristics and kinetics of corn stover and its digestate by TG-FTIR. BioResources 12, 8240–
 8254. https://doi.org/10.15376/biores.12.4.8240-8254